# UNIVERSITY of MISSOURI

### DEPARTMENT OF VETERINARY MEDICINE & SURGERY

COLLEGE OF VETERINARY MEDICINE

#### Genetic Notes on the Burmese Cat Breed

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The Feline Genetics Laboratory, formerly of the University of California – Davis (UC Davis), now at the University of Missouri, has had long associations with Burmese breeders and veterinary researchers to define and identify the genetic mutations that cause various maladies in the Burmese breed. Many published studies (several listed below) have demonstrated that the Burmese breed has many genetic conditions, although the prevalence of these conditions is variable between countries. The Burmese in the USA are historically known for the autosomal recessive craniofacial defect that developed from the Contemporary lines of USA Burmese. Burmese carrying this condition have been identified in the UK, Europe and Australia due to the importation and use of the Contemporary lines. In addition, any other breed that has outcrossed to Contemporary Burmese has risk of the craniofacial defect. Recently, American Burmese breeders may be pursuing an organized project for heart disease.

In contrast, non - American Burmese are afflicted with several other conditions including GM2 gangliosidosis, hypokalemia, oral facial pain, diabetes, high triglycerides, a midline closure defect, and dermatosparaxis (stretchy skin, cutaneous asthenia, Ehlors-Danlos Syndrome). The genetic mutations for hypokalemia and GM2 gangliosidosis have been identified and like the craniofacial defect, the genetic test is available to assist breed management. Genetics studies regarding oral facial pain, diabetes, high triglycerides, flat-chested kittens, and dermatosparaxis are underway by our laboratory, in collaboration with other researchers, and independently by other researchers.

Our genetic studies have also included examining the population genetics of the different cat breeds and worldwide populations. In 2007, we published the first breed study that demonstrated that Singapura, Burmese and Birman had the lowest genetic diversity of the cat breeds we examined. A study between Singapura breeders and the UC Davis Veterinary Genetics Laboratory (VGL) supported our findings. Our recent studies demonstrate that the non-USA Burmese has as low genetic variation as the USA lines of Burmese. We strongly recommended outcrossing for all the Burmese breeds, including USA and non-USA Burmese.

Based on genetics, the Burmese breed originated from cats of Southeast Asia. Our genetic studies group cats from Thailand, Cambodia, Vietnam, Brunei and the Philippines as Southeast Asia cats. Historically, Burmese are considered to have origins from Thailand. Thus, random bred cats from Thailand and perhaps other regions of Southeast Asia are recommended as strong genetic candidates for outcrossing. Genetic tests for colors and health conditions can assist the selection of



cats that may be the most advantageous for outcrossing. Other breeds with Southeast origins would also be of value to the Burmese outcrossing program, including Singapura, Bombay, and Burmese from any country. Burmilla and Asian have been crossed with Persian cats, which have western origins, thus, the use of these recent crossbred breed cats would add significant diversity, but of a different "racial" origins. Other Southeast Asian breeds include Siamese (and its breed derivatives), Korat, and Birman.

Because of our various genetic projects that include Burmese from different countries, we have been able to compare the genetics of the Burmese breed groups regionally. USA Burmese have high homozygosity (0.38), implying low genetic diversity (See attached report). The non-USA Burmese, which is a mixture of UK and Australian Burmese also have low diversity and high homozygosity (0.41), as compared to random bred cats (0.04) and other breeds. When we examine the Burmese diversity by plotting the genetic differences (termed multi-dimensional scaling (MDS), we find that the USA and non-USA Burmese do not cluster together and form independent genetic groups. The USA cats have a larger spread to their cluster as it includes Contemporary and Traditional lines of Burmese. Thus, Burmese breeders may consider crossing the Burmese cats from different countries to help improve genetic diversity.

Other genetic conditions in Southeast Asian cats include different forms of gangliosidosis in Korat and Siamese cats and pyruvate kinase deficiency in Singapura. Some Birman lines may have a susceptibility to FIP.

Overall, we have more genetic projects concerning Burmese than for any other breed and our various genetic studies have repeatedly demonstrated that Burmese, regardless of country, have amongst the lowest genetic diversity of all cat breeds. An organized and multi-faceted outcrossing program for the breed is highly recommended which includes the use of genetic testing and outcrossing to different populations and breeds.

Best regards,

Leslie A. Lyons, PhD

#### **Text Information References**

- 1: Kurushima JD, Lipinski MJ, Gandolfi B, Froenicke L, Grahn JC, Grahn RA, Lyons LA. Variation of cats under domestication: genetic assignment of domestic cats to breeds and worldwide random-bred populations. Anim Genet. 2013 Jun;44(3):311-24. doi: 10.1111/age.12008. Epub 2012 Nov 22. PubMed PMID: 23171373; PubMed Central PMCID: PMC3594446.
- 2: Lipinski MJ, Froenicke L, Baysac KC, Billings NC, Leutenegger CM, Levy AM, Longeri M, Niini T, Ozpinar H, Slater MR, Pedersen NC, Lyons LA. The ascent of cat breeds: genetic evaluations of breeds and worldwide random-bred populations. Genomics. 2008 Jan;91(1):12-21. Epub 2007 Dec 3. PubMed PMID: 18060738; PubMed Central PMCID: PMC2267438.
- 3: Lyons LA, Erdman CA, Grahn RA, Hamilton MJ, Carter MJ, Helps CR, Alhaddad H, Gandolfi B. Aristaless-Like Homeobox protein 1 (ALX1) variant associated with craniofacial structure and frontonasal dysplasia in Burmese cats. Dev Biol. 2016 Jan 15;409(2):451-8. doi: 10.1016/j.ydbio.2015.11.015. Epub 2015 Dec 2. PubMed PMID: 26610632; PubMed Central PMCID: PMC4724490.
- 4: Malik R, Musca FJ, Gunew MN, Menrath VH, Simpson C, Culvenor J, Grahn RA, Helps C, Lyons LA, Gandolfi B. Periodic hypokalaemic polymyopathy in Burmese and closely related cats: a review including the latest genetic data. J Feline Med Surg. 2015 May;17(5):417-26. doi: 10.1177/1098612X15581135. Review. PubMed PMID: 25896241.
- 5: O'Leary CA, Duffy DL, Gething MA, McGuckin C, Rand JS. Investigation of diabetes mellitus in Burmese cats as an inherited trait: a preliminary study. N Z Vet J. 2013 Nov;61(6):354-8. doi: 10.1080/00480169.2013.817295. Epub 2013 Aug 5. PubMed PMID: 23909918.
- 6: Alhaddad H, Khan R, Grahn RA, Gandolfi B, Mullikin JC, Cole SA, Gruffydd-Jones TJ, Häggström J, Lohi H, Longeri M, Lyons LA. Extent of linkage disequilibrium in the domestic cat, Felis silvestris catus, and its breeds. PLoS One. 2013;8(1):e53537. doi: 10.1371/journal.pone.0053537. Epub 2013 Jan 7. PubMed PMID: 23308248; PubMed Central PMCID: PMC3538540.
- 7: Gandolfi B, Gruffydd-Jones TJ, Malik R, Cortes A, Jones BR, Helps CR, Prinzenberg EM, Erhardt G, Lyons LA. First WNK4-hypokalemia animal model identified by genome-wide association in Burmese cats. PLoS One. 2012;7(12):e53173. doi: 10.1371/journal.pone.0053173. Epub 2012 Dec 28. PubMed PMID: 23285264; PubMed Central PMCID: PMC3532348.
- 8: Rusbridge C, Heath S, Gunn-Moore DA, Knowler SP, Johnston N, McFadyen AK. Feline orofacial pain syndrome (FOPS): a retrospective study of 113 cases. J Feline Med Surg. 2010 Jun;12(6):498-508. doi: 10.1016/j.jfms.2010.03.005. Epub 2010 May 6. PubMed PMID: 20451434.

- 9: Kluger EK, Hardman C, Govendir M, Baral RM, Sullivan DR, Snow D, Malik R. Triglyceride response following an oral fat tolerance test in Burmese cats, other pedigree cats and domestic crossbred cats. J Feline Med Surg. 2009 Feb;11(2):82-90. doi: 10.1016/j.jfms.2008.05.005. Epub 2008 Jul 30. PubMed PMID: 18667349.
- 10: Lederer R, Rand JS, Jonsson NN, Hughes IP, Morton JM. Frequency of feline diabetes mellitus and breed predisposition in domestic cats in Australia. Vet J. 2009 Feb;179(2):254-8. Epub 2007 Dec 21. PubMed PMID: 18155627.
- 11: McCann TM, Simpson KE, Shaw DJ, Butt JA, Gunn-Moore DA. Feline diabetes mellitus in the UK: the prevalence within an insured cat population and a questionnaire-based putative risk factor analysis. J Feline Med Surg. 2007 Aug;9(4):289-99. Epub 2007 Mar 27. PubMed PMID: 17392005.
- 12: Lantinga E, Kooistra HS, van Nes JJ. [Periodic muscle weakness and cervical ventroflexion caused by hypokalemia in a Burmese cat]. Tijdschr Diergeneeskd. 1998 Jul 15-Aug 1;123(14-15):435-7. Dutch. PubMed PMID: 9700861.
- 13: Rand JS, Bobbermien LM, Hendrikz JK, Copland M. Over representation of Burmese cats with diabetes mellitus. Aust Vet J. 1997 Jun;75(6):402-5. PubMed PMID: 9247686.
- 14: Sponenberg DP, Graf-Webster E. Hereditary meningoencephalocele in Burmese cats. J Hered. 1986 Jan-Feb;77(1):60. PubMed PMID: 2937834.
- 15: Hansen N, Foster SF, Burrows AK, Mackie J, Malik R. Cutaneous asthenia (Ehlers-Danlos-like syndrome) of Burmese cats. J Feline Med Surg. 2015 Nov;17(11):954-63. doi: 10.1177/1098612X15610683. PubMed PMID: 26486982.
- 16: Bradbury AM, Morrison NE, Hwang M, Cox NR, Baker HJ, Martin DR. Neurodegenerative lysosomal storage disease in European Burmese cats with hexosaminidase beta-subunit deficiency. Mol Genet Metab. 2009 May;97(1):53-9. doi: 10.1016/j.ymgme.2009.01.003. Epub 2009 Feb 23. PubMed PMID: 19231264.
- 17: Lyons LA, Imes DL, Rah HC, Grahn RA. Tyrosinase mutations associated with Siamese and Burmese patterns in the domestic cat (Felis catus). Anim Genet. 2005 Apr;36(2):119-26. PubMed PMID: 15771720.

#### **Table References**

- 1. Eizirik E, Yuhki N, Johnson WE, et al. **Molecular genetics and evolution of melanism** in the cat family. *Curr Biol* 2003; 13: 448-453.
- 2. Gershony LC, Penedo MC, Davis BW, et al. Who's behind that mask and cape? The Asian leopard cat's Agouti (ASIP) allele likely affects coat colour phenotype in the Bengal cat breed. *Anim Genet* 2014; 45: 893-897.

- 3. Lyons LA, Foe IT, Rah HC, et al. **Chocolate coated cats: TYRP1 mutations for brown color in domestic cats**. *Mamm Genome* 2005; 16: 356-366.
- 4. Schmidt-Kuntzel A, Eizirik E, O'Brien SJ, et al. *Tyrosinase* and *tyrosinase* related protein 1 alleles specify domestic cat coat color phenotypes of the albino and *Brown* loci. *J Hered* 2005; 96: 289-301.
- 5. Imes DL, Geary LA, Grahn RA, et al. **Albinism in the domestic cat (Felis catus) is associated with a tyrosinase (TYR) mutation**. *Anim Genet* 2006; 37: 175-178.
- 6. Lyons LA, Imes DL, Rah HC, et al. **Tyrosinase mutations associated with Siamese and Burmese patterns in the domestic cat (Felis catus)**. *Animal Genet* 2005; 36: 119-126.
- 7. Ishida Y, David VA, Eizirik E, et al. A homozygous single-base deletion in MLPH causes the dilute coat color phenotype in the domestic cat. *Genomics* 2006.
- 8. Peterschmitt M, Grain F, Arnaud B, et al. **Mutation in the melanocortin 1 receptor is associated with amber colour in the Norwegian Forest Cat**. *Anim Genet* 2009; 40: 547-552.
- 9. Lyons LA, Creighton EK, Alhaddad H, et al. Whole Genome Sequencing Identifies an AIPL1 Variant in Persian Cats as a New Model for Leber's Congenital Amaurosis. *BMC Genome* 2016; (In press)).
- 10. Drogemuller C, Rufenacht S, Wichert B, et al. **Mutations within the FGF5 gene are associated with hair length in cats**. *Anim Genet* 2007; 38: 218-221.
- 11. Kehler JS, David VA, Schaffer AA, et al. Four independent mutations in the feline fibroblast growth factor 5 gene determine the long-haired phenotype in domestic cats. *J Hered* 2007; 98: 555-566.
- 12. Kaelin CB, Xu X, Hong LZ, et al. **Specifying and sustaining pigmentation patterns in domestic and wild cats**. *Science* 2012; 337: 1536-1541.
- 13. David VA, Menotti-Raymond M, Wallace AC, et al. **Endogenous retrovirus insertion in the** *KIT* **oncogene determines** *White* **and white** *Spotting* **in domestic cats**. *G3* 2014; 4: 1881-1891.
- 14. Bighignoli B, Niini T, Grahn RA, et al. **Cytidine monophospho-N-acetylneuraminic acid hydroxylase (CMAH) mutations associated with the domestic cat AB blood group.** *BMC Genet* 2007; 8: 27.
- 15. Gandolfi B, Grahn RA, Gustafson N, et al. **Type it! The genetic characterization of CMAH in Ragdoll blood type AB**. *BMC veterinary research* 2016; (Submitted).
- 16. Lyons LA, Erdman CA, Grahn RA, et al. **Aristaless-Like Homeobox protein 1 (ALX1)** variant associated with craniofacial structure and frontonasal dysplasia in Burmese cats. *Dev Biol* 2015
- 17. De Maria R, Divari S, Bo S, et al. **Beta-galactosidase deficiency in a Korat cat: a new form of feline GM1-gangliosidosis.** *Acta Neuropathol (Berl)* 1998; 96: 307-314.
- 18. Bradbury AM, Morrison NE, Hwang M, et al. **Neurodegenerative lysosomal storage disease in European Burmese cats with hexosaminidase beta-subunit deficiency**. *Mol Genet Metab* 2009; 97: 53-59.
- 19. Muldoon LL, Neuwelt EA, Pagel MA, et al. **Characterization of the molecular defect in a feline model for type II GM2-gangliosidosis (Sandhoff disease)**. *Am J Pathol* 1994; 144: 1109-1118.
- 20. Gandolfi B, Gruffydd-Jones TJ, Malik R, et al. **First WNK4-hypokalemia animal model identified by genome-wide association in Burmese cats**. *PloS one* 2012; 7: e53173.
- 21. Grahn RA, Grahn JC, Penedo MC, et al. **Erythrocyte pyruvate kinase deficiency mutation identified in multiple breeds of domestic cats**. *BMC veterinary research* 2012; 8: 207.

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Table 1 The phenotypic traits of the domestic cat – special interest for Burmese

Disease / Trait (alleles) OMIA Entry	MOI <sup>‡</sup>	Phenotype	Gene	Gene Name	Mutation
Agouti (A <sup>+</sup> , a, A <sup>PDE</sup> ) <sup>1, 2</sup> 000201-9685	AR	Banded fur to solid	ASIP	Agouti-signaling protein	c.122_123delCA; Pbe haplotype
Brown (B <sup>+</sup> , b, b') <sup>3, 4</sup> 001249-9685	AR	Brown, light brown color variants	TYRP1	Tyrosinase related protein	b = -5IVS6, b <sup>l</sup> = c.298C>T
Color (C <sup>+</sup> , C <sup>b</sup> , C <sup>s</sup> , c) <sup>4-6</sup> 000202-9685	AR	Burmese, Siamese color pattern, full albino	TYR	Tyrosinase	$c^{b} = c.715G>T, c^{s} = c.940G>A, c$ = c.975delC
Dilution (D <sup>+</sup> , d) <sup>'</sup> 000206-9685	AR	Black to grey / blue, Orange to cream	MLPH	Melanophilin	c.83delT
Extension ( $E^{+}$ , e, e') – Amber <sup>8</sup> 001199-9685	AR	Brown/red color variant	MC1R	Melanocortin receptor 1	c.250G>A; c.439TCT
Japanese Bobtail (J, j <sup>+</sup> ) <sup>9</sup>	AD	Kinked tail	HES7	Hairy and Enhancer of Split family, transcription factor 7	c.5A>G
Longhair (L <sup>+</sup> , I) <sup>10, 11</sup> 000439-9685	AR	Long fur	FGF5	Fibroblast growth factor 5	c.356_367insT, c.406C>T, c.474delT, c.475A>C
Orange (O, o <sup>+</sup> )	X linked	Change in pigment hue	unknown	unknown	unknown
Tabby(T <sup>M</sup> , t <sup>b</sup> ) <sup>12</sup> 001429-9685	AR	Blotched/classic pattern	TAQPEP	Transmembrane aminopeptidase Q	S59X, T139N, D228N, W841X
Ticked (T <sup>a</sup> , t) 001484-9685	AD	No Tabby pattern	unknown	unknown	unknown
White (W, w <sup>+</sup> ) <sup>13</sup> 000209-9685	AD	Loss of pigmentation	KIT	KIT	FERV1 LTR ins
Wide-band	AR?	Length of pheomelanin band in hair	unknown	unknown	unknown

<sup>‡</sup> Mode of inheritance of the non-wild type variant. A "+" implies the wild type allele when known. In reference to the mutant allele, AD implies autosomal dominant, AR implies autosomal recessive, co-D implies co-dominant. OMIA: Online Mendelian Inheritance in Animals (http://omia.angis.org.au/home/) entries provides links to citations and clinical descriptions of the phenotypes and the diseases. Presented citations are for the causative variant discovery.

Table 2 Inherited diseases of domestic cats for which a commercial DNA test is available - special interest for Burmese

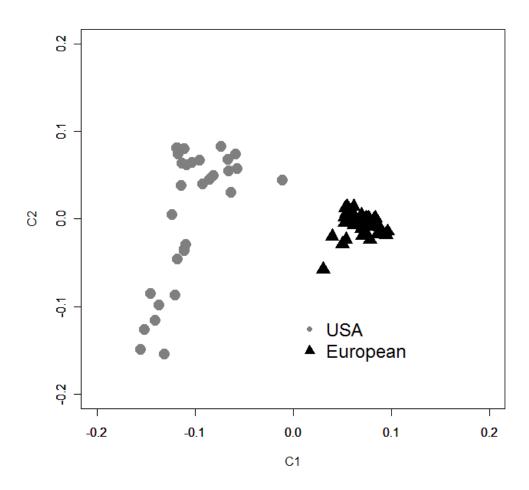
Disease / Trait (alleles) OMIA					
Entry	MOI <sup>‡</sup>	Phenotype	Gene	Gene Name	Mutation
AB Blood Type (A <sup>+</sup> , AB, b) <sup>14,</sup> 15 000119-9685	AR	Determines Type B	СМАН	cytidine monophospho-N- acetylneuraminic acid hydroxylase	c.1del-53_70, c.139G>A
Craniofacial Defect <sup>16</sup>	AR	Craniofacial Defect	ALX1	Aristaless-Like Homeobox 1	c.496delCTCTCAGGACTG
Gangliosidosis 1 <sup>17</sup> 000402- 9685	AR	Lipid storage disorder (GM1)	GLB1	Galactosidase, beta 1	c.1457G>C
Gangliosidosis 2 <sup>18</sup> 01462-0985	AR	Lipid storage disorder (GM2)	HEXB	Hexominidase B	c.1356del-1_8, c.1356_1362delGTTCTCA
Gangliosidosis 2 <sup>19</sup> 01462-0985	AR	Lipid storage disorder (GM2)	HEXB	Hexominidase B	c.39delC
Hypokalemia <sup>20</sup> 001759-9685	AR	Potassium deficiency (HK)	WNK4	WNK lysine deficient protein kinase 4	c.2899C>T
Pyruvate Kinase Def. <sup>21</sup> 000844-9685	AR	Hemopathy (PK Deficiency)	PKLR	pyruvate kinase, liver, RBC	c.693+304G>A

<sup>‡</sup> Mode of inheritance of the non-wild type variant. Not all transcripts for a given gene may have been discovered or well documented in the cat, mutations presented as interpreted from original publication. A "+" implies the wild type allele when known. In reference to the mutant allele, AD implies autosomal dominant, AR implies autosomal recessive, co-D implies co-dominant. OMIA: Online Mendelian Inheritance in Animals (http://omia.angis.org.au/home/) entries provides links to citations and clinical descriptions of the phenotypes and the diseases. Presented citations are for the causative variant discovery.

<u>Inbreeding coefficient</u> based on the observed versus expected number of homozygous genotypes in each sample.

USA Burmese 0.38
European Burmese 0.41
Random bred 0.04
LaPerm 0.07
Selkirk rex 0.15
Bengal 0.19
American curl 0.11
Japanese bobtail 0.14

### MDS American vs European Burmese:



European Burmese cluster very tightly, while American Burmese are more interspersed, this is an indication of higher genetic diversity within the American Burmese.